

1 **Supplemental Material for**

2 **Sorption of lincomycin by manure-derived biochars from water**

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22 **Supplemental Methods**

23 ***Zeta Potential Measurement***

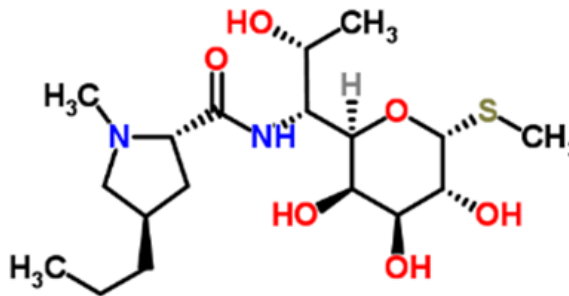
24 Zeta potential of biochar particles was measured with the Malvern Zetasizer Nano-ZS
25 equipped with a MPT-2 autotitrator (Malvern Instruments, Worcestershire, UK). To generate
26 biochar colloid suspension for isoelectric point (pH_{IEP}) measurement, 10 mg biochar (particle size
27 $< 75 \mu\text{m}$) was mixed with 50 mL DI water in a polyethylene centrifuge tube and then sonicated for
28 30 min. After sonication, top 10 mL of biochar suspension was withdrawn and then titrated with
29 0.1 M HCl or NaOH titrant from pH 10 to 2 using the autotitrator, and the corresponding zeta
30 potential at each pH was measured by the Zetasizer Nano-ZS. The pH_{IEP} value was determined at
31 the pH where the zeta potential is zero. Additionally, zeta potential for biochar suspensions of 1-
32 day or 180-day water exposure was also determined by the zetasizer.

33 ***LC-MS/MS Analytical Procedure***

34 Lincomycin concentrations in the solution were determined by a Shimadzu Prominence high-
35 performance liquid chromatograph coupled to an Applied Biosystems Sciex 3200 triple
36 quadrupole mass spectrometer (LC-MS/MS). A Gemini 5u C18 110A 50 \times 2.00 mm 5 μm column
37 was used. The mobile phase consisted of water (A) and 1:1 (v/v) acetonitrile-methanol mixture (B)
38 with A and B both containing 0.3% formic acid. Gradient conditions were 0 % to 40 % B in 0 to 1
39 minute, 40 % to 70% B in 1 to 2 minutes, 70 % – 80 % B in 2 to 3 minutes, 80 % to 100 % B in 3
40 to 3.5 minutes, and held for 0.5 minutes at a flow rate of 0.35 mL/min. Injection volume was 10
41 μL . The tandem quadrupole MS was used with an electrospray ionization (ESI) and positive ion
42 mode. Lincomycin was detected and quantified using a multiple reaction monitoring mode with a
43 precursor/product transition of 407.2/126.2. The retention time and instrument detection limit of
44 lincomycin was 2.37 min and 0.2 pg.

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46 Table S1. Chemical and physical properties of lincomycin[†]

Properties	Lincomycin
Molecular structure [‡]	 <p>The image shows the chemical structure of Lincomycin. It consists of a 2-methyl-5-propylpyrrolidine ring connected via an amide bond to a 2,6-dimethyl-4-(methylsulfanyl)tetrahydropyran ring. The tetrahydropyran ring has three hydroxyl groups at the 2, 3, and 4 positions. Stereochemistry is indicated with wedges and dashes.</p>
Molecular Formula [§]	C ₁₈ H ₃₄ N ₂ O ₆ S
Molecular weight [§]	406.54
pKa [§]	7.6
log K _{ow} [§]	0.20
Water solubility [§]	927 mg L ⁻¹ at 25 °C

47 [†] pKa: dissociation constant, K_{ow}: octanol/water partition coefficient; [‡] Data from ChemSpider
 48 (<http://www.chemspider.com/>); [§] Data from TOXNET (<http://www.toxnet.nlm.nih.gov/>)

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50 Table S2. Fitted parameters of pseudo-first-order, pseudo-second-order, and intraparticle
 51 diffusion models for long-term sorption kinetics of lincomycin on manure-derived biochars.

Biochar	Pseudo-first-order			
	q_e ($\mu\text{g}\cdot\text{g}^{-1}$)	k_1 (day^{-1})	R^2	RMSE
BM600	937	3.11×10^{-2}	0.837	175
DM600	855	3.37×10^{-2}	0.951	187
AM600	851	3.13×10^{-2}	0.867	203
PM600	860	3.07×10^{-2}	0.837	197
Biochar	Pseudo-second-order			
	q_e ($\mu\text{g g}^{-1}$)	k_2 ($\mu\text{g g}^{-1} \text{day}^{-1}$)	R^2	RMSE
BM600	856	9.91×10^{-5}	0.916	144
DM600	990	1.46×10^{-4}	0.987	124
AM600	907	1.40×10^{-4}	0.959	142
PM600	852	1.13×10^{-4}	0.927	169
Biochar	Intraparticle diffusion			
	C ($\mu\text{g}\cdot\text{g}^{-1}$)	k_i ($\mu\text{g g}^{-0.5} \text{day}^{-0.5}$)	R^2	RMSE
BM600	160	52.3	0.973	34.0
DM600	212	66.2	0.965	49.7
AM600	228	54.6	0.963	42.1
PM600	216	47.0	0.972	31.2

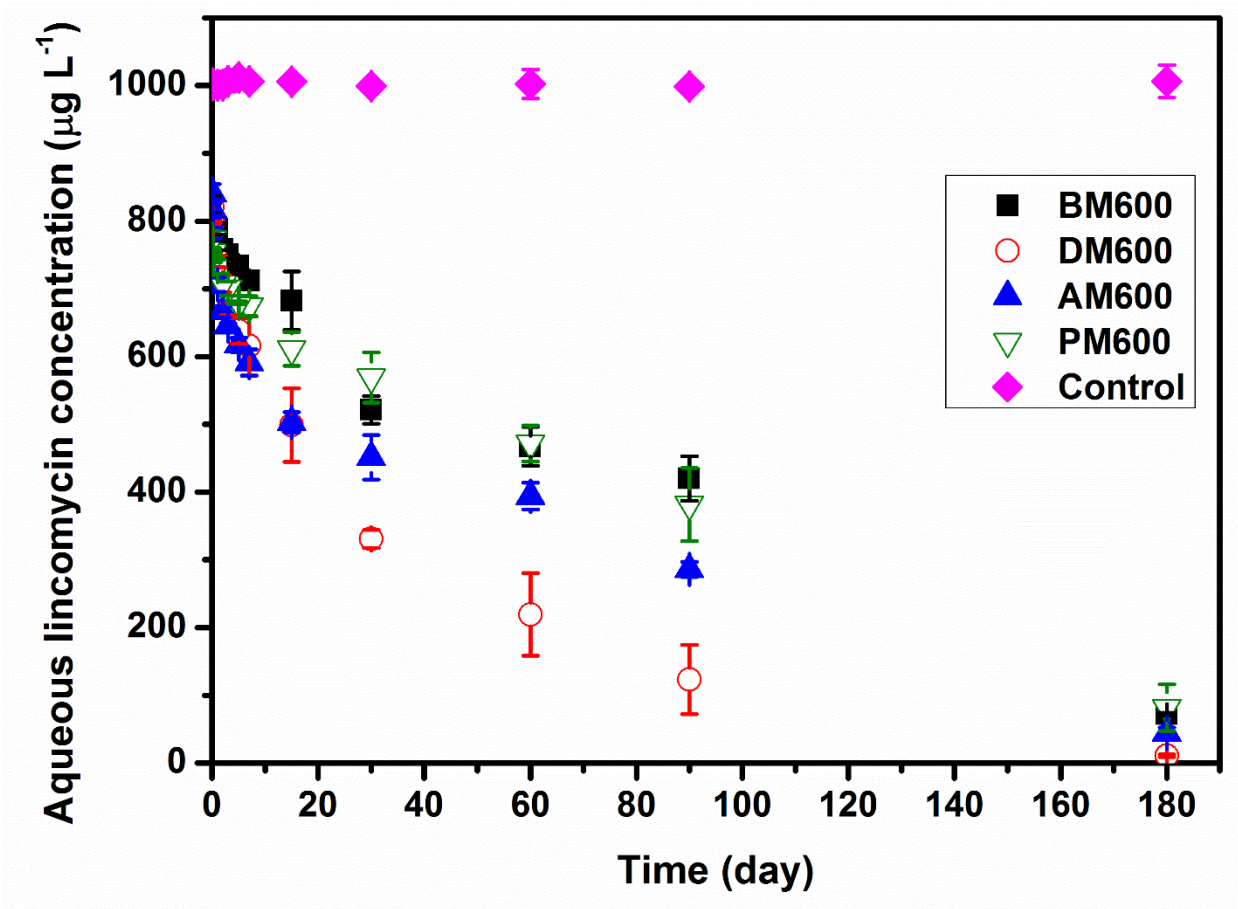
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53 Table S3. Fitted parameters of Langmuir and Freundlich equations for lincomycin sorption on
 54 manure-derived biochars.

Biochar	Langmuir				Freundlich			
	q_{\max} ($\mu\text{g g}^{-1}$)	K_L ($\text{L } \mu\text{g}^{-1}$)	R^2	RMSE	n^{-1}	K_F ($\mu\text{g}^{(n-1)} \text{g}^{-1} \text{L}^n$)	R^2	RMSE
BM600 pH 6.6	555	1.29×10^{-2}	0.99	15	0.47	28.5	0.98	34
BM600 pH 9.9	299	7.87×10^{-3}	1.00	5.2	0.44	15.4	0.96	18
DM600 pH 6.5	605	1.40×10^{-2}	0.99	19	0.48	30.7	0.97	38
DM600 pH 10.0	372	5.68×10^{-3}	0.97	17	0.48	13.4	0.97	13
AM600 pH 6.9	697	6.18×10^{-3}	1.00	9.3	0.61	13.7	0.99	29
AM600 pH 10.0	436	4.58×10^{-3}	0.97	18	0.55	10.1	0.95	19
PM600 pH 7.3	576	2.68×10^{-3}	0.96	15	0.71	4.29	0.96	29
PM600 pH 10.4	424	3.41×10^{-3}	0.98	12	0.59	6.83	0.98	11

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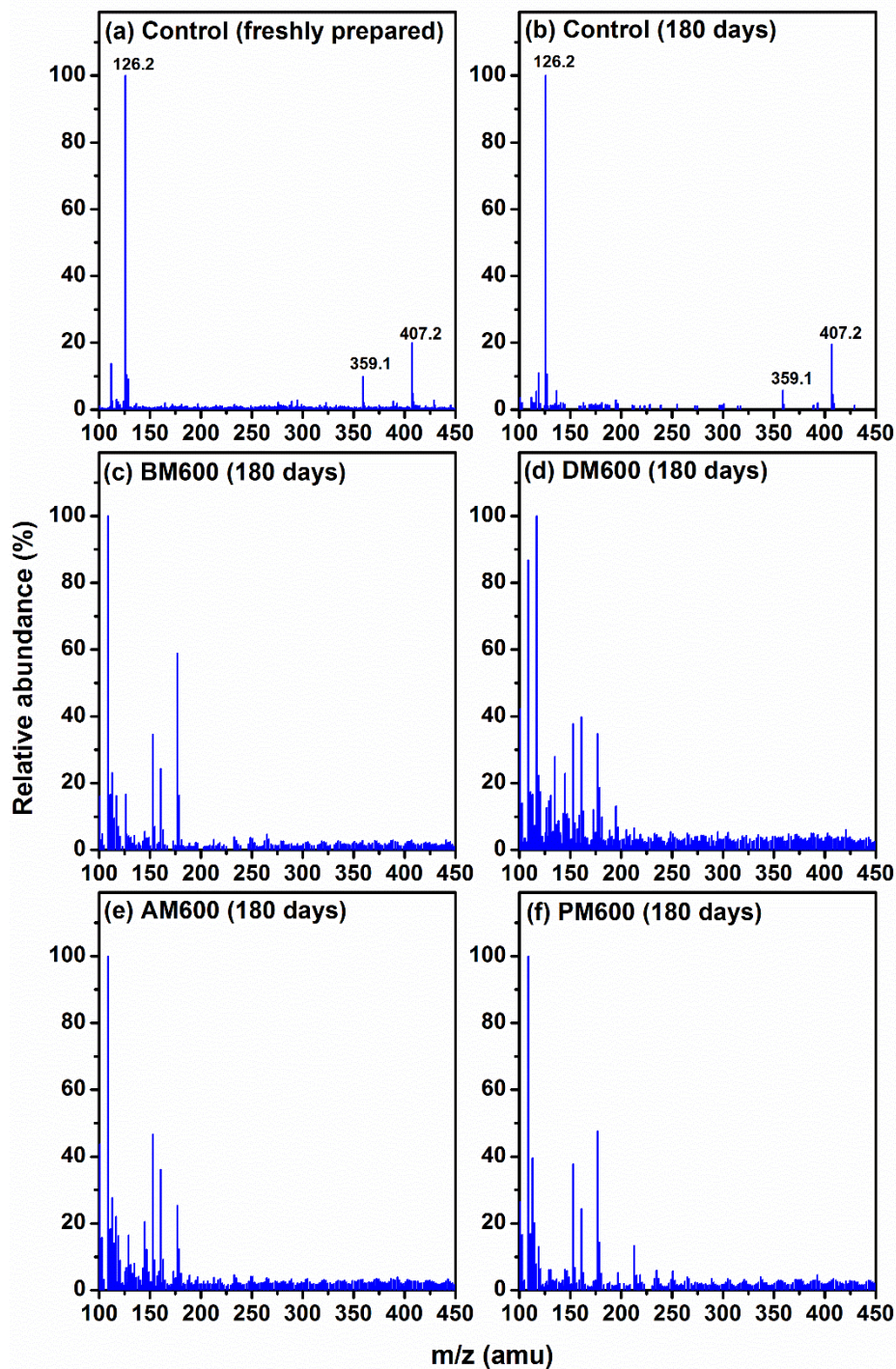


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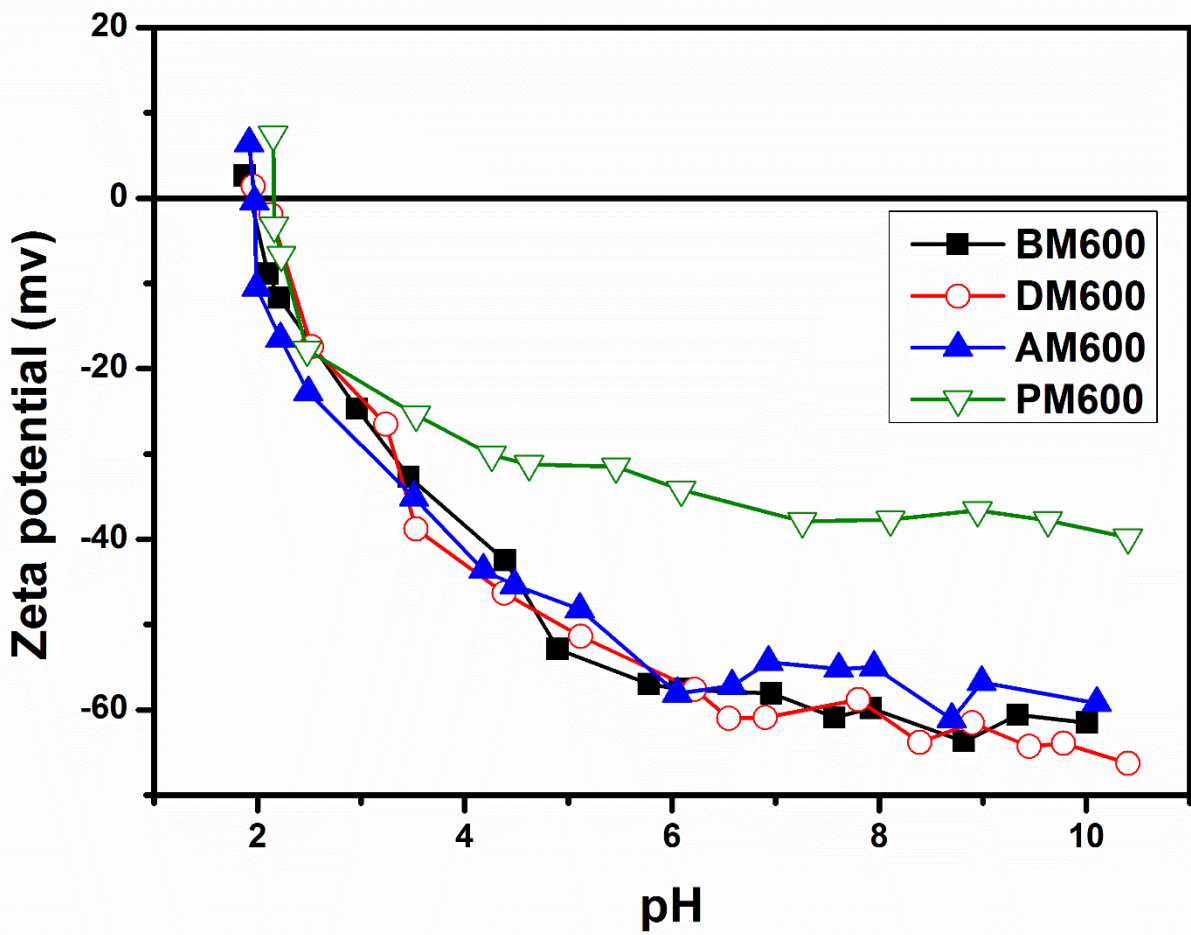
58 Fig S1. Lincomycin concentration versus time for lincomycin sorption kinetics experiments.

59 Control was the biochar-free lincomycin solution.

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 62 Fig S2. Precursor ion scan spectra of (a) Control (freshly prepared), (b) Control (180 days), (c)
 63 BM600 (180 days), (d) DM600 (180 days), (e) AM600 (180 days), and (f) PM600 (180days).
 64 Control was the biochar-free lincomycin solution. No degradation candidates of lincomycin was
 65 detected in long-term kinetics samples (b, d, f).



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67 Fig S3. Zeta potential of manure-derived biochars as a function of solution pH.